

Evaluation of soil microfungi as biological control agents against ascarid eggs

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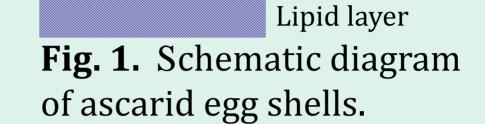
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Introduction

Materials and methods

- Thick-shelled ascarid eggs may remain infective in the environment for several years, thus posing a prolonged risk of transmission to animals or humans
- → Aim: to evaluate if microfungi (*Pochonia chlamydosporia* and *Paecilomyces lilacinus*) could reduce the viability of *Ascaridia galli, Toxocara canis* and *Ascaris suum* eggs under laboratory conditions





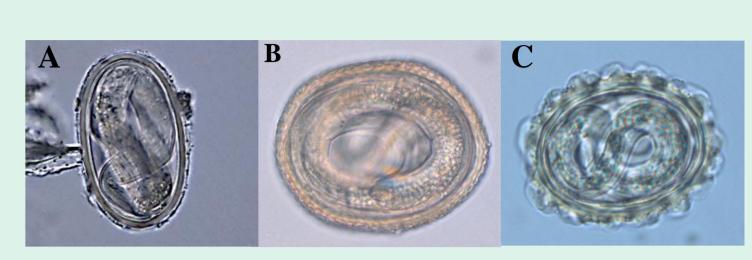


Fig. 2. Embryonated ascarid eggs: A. *Ascaridia* galli, B. *Toxocara canis*, C. *Ascaris suum*.

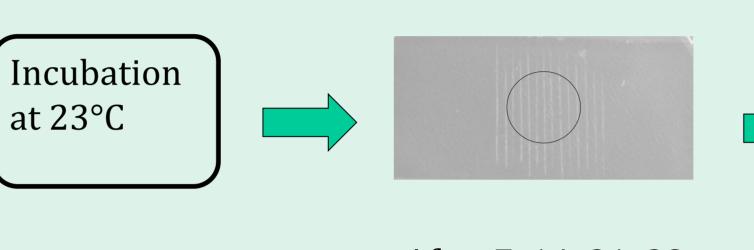
4.9 cm² premarked area P. chlamydosporia (biotype 10) or P. lilacinus (strain 251)

P. chlamydosporia (biotype 10) or P. lilacinus (strain 251) inoculated on 2% water agar (1.5 mm thick) covering a glass slide inside a Petri dish

Control Petri dishes were not treated with any fungus

For each parasite species: approx. 150 fresh eggs added to the premarked area on Petri dishes with and without

Approx. number of eggs counted *in* situ using a stereomicroscope (day 0 count)



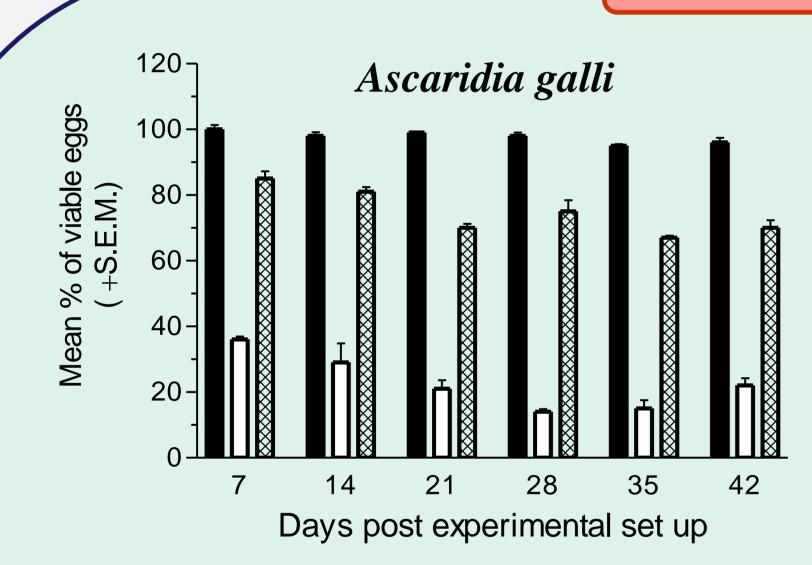
After 7, 14, 21, 28, 35 and 42 days, the glass slide was released from the Petri dish (n=3 for each time point and treatment)

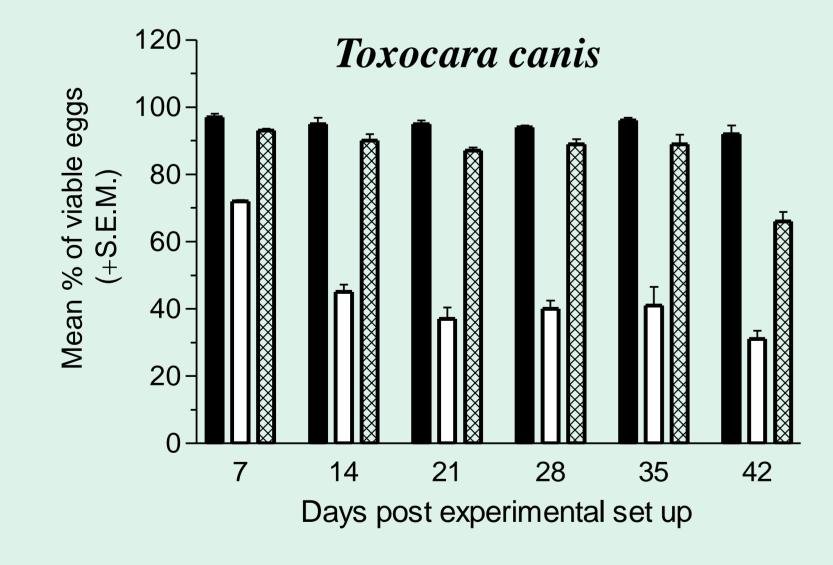
Egg viability (= ability to embryonate) examined using compound and differential interference contrast microscopes

Percentage of viable eggs calculated in relation to day 0 count

Results

fungi





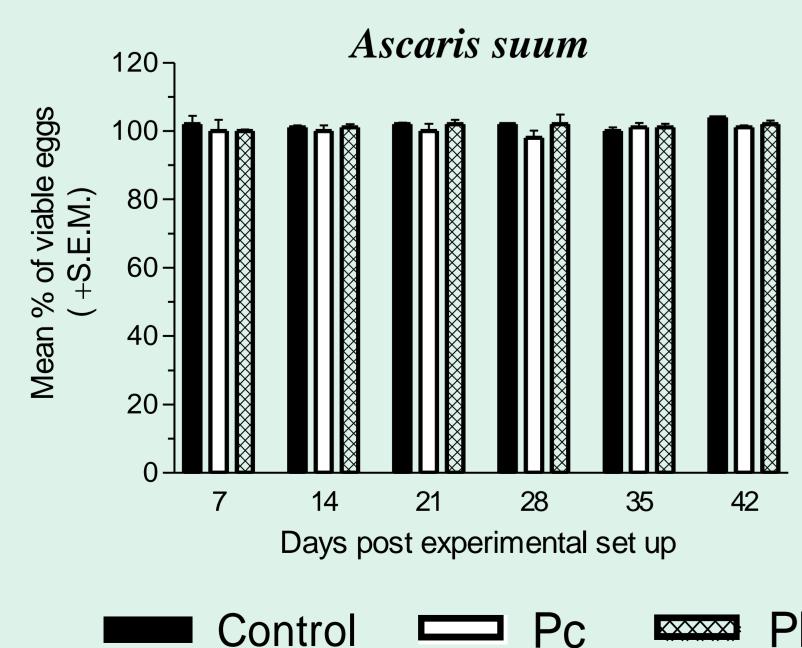


Fig. 3. Mean percentages (n=3, +S.E.M) of viable *A. galli, T. canis* and *A. suum* eggs on control, or a fungus (*P. chlamydosporia* (Pc) or *P. lilacinus* (Pl)) treated plates.

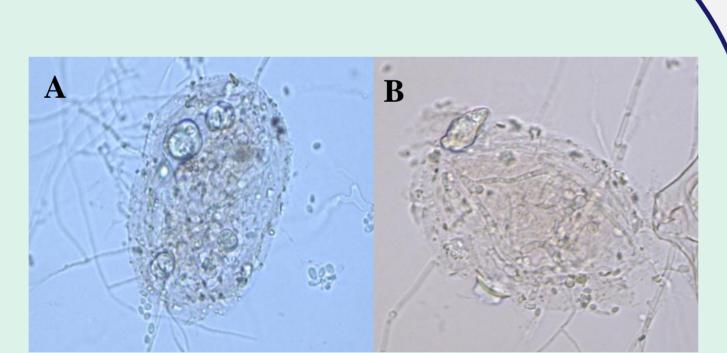


Fig. 4. *A. galli* eggs destroyed by *P. chlamydosporia* (A, B). Both shell and contents are degenerated.

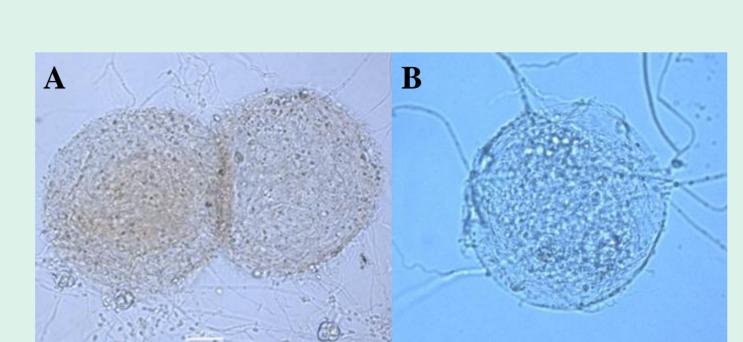


Fig. 5. *T. canis* eggs destroyed by *P. chlamydosporia* (A) and *P. lilacinus* (B). Both shell and contents are degenerated.

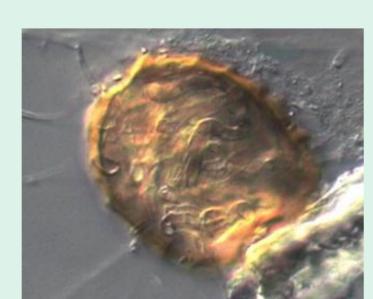


Fig. 6. *A. suum* egg destroyed by *P. chlamydosporia*. Most of the shell is intact whereas the content is degenerated and colonized by hyphae.

Discussion and conclusion

- *P. chlamydosporia* had a large negative impact on viability of *A. galli* and *T. canis* eggs whereas *P. lilacinus* had only a limited effect → interspecies differences in the ability of fungi to destroy eggs
- *A. suum* eggs more resistant than *A. galli* and *T. canis* eggs to both fungi → the thick uterine layer in *A. suum* eggs is perhaps important in protecting against the microfungi
- Shells and contents of *A. galli* and *T. canis* eggs were completely degraded → enzymatic degradation of egg shell protein and chitin (Khan et al., 2004; Zhang et al., 2009) probably the primary mechanism of both fungi
- → *P. chlamydosporia* may be a potential biological control agent against *A. galli* and *T. canis* eggs in the environment

Acknowledgements

Rothamsted Research (UK) and Prophyta (Germany) are thanked for kindly providing the fungal isolates *Pochonia chlamydosporia* biotype 10 and *Paecilomyces lilacinus* strain 251, respectively. The study was funded by the Green Development and Demonstration Programme (GUDP), the Danish AgriFish Agency, Danish Ministry for Food, Agriculture and Fisheries.

References

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Zhang, C., Wu, X., Cai, X., 2009. Effect of chitinases produced by *Pochonia chlamydosporia* on egg-hatching of *Meloidogyne incognita*. Sci. Agri. Sin. 42, 3509-3515.