Effects of vertical distribution of soil inorganic nitrogen on root growth and subsequent nitrogen uptake by field vegetable crops

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Abstract

Information is needed about root growth and N uptake of crops under different soil conditions to increase nitrogen use efficiency in horticultural production. The purpose of this study was to investigate if differences in vertical distribution of soil nitrogen (N_{inorg}) affected root growth and N uptake of a variety of horticultural crops. Two field experiments were performed each over 2 years with shallow or deep placement of soil N_{inorg} obtained by management of cover crops. Vegetable crops of leek, potato, Chinese cabbage, beetroot, summer squash and white cabbage reached root depths of 0.5, 0.7, 1.3, 1.9, 1.9 and more than 2.4 m, respectively, at harvest, and showed rates of root depth penetration from 0.2 to 1.5 mm day⁻¹ °C⁻¹. Shallow placement of soil N_{inorg} resulted in greater N uptake in the shallow-rooted leek and potato. Deep placement of soil N_{inorg} resulted in greater rates of root depth penetration in the deep-rooted Chinese cabbage, summer squash and white cabbage, which increased their depth by 0.2–0.4 m. The root frequency was decreased in shallow soil layers (white cabbage) and increased in deep soil layers (Chinese cabbage, summer squash and white cabbage). The influence of vertical distribution of soil N_{inorg} on root distribution and capacity for depletion of soil N_{inorg} was much less than the effect of inherent differences between species. Thus, knowledge about differences in root growth between species should be used when designing crop rotations with high N use efficiency.

Keywords: Deep roots, horticulture, nitrate, minirhizotron, root density, soil nitrogen, subsoil

Introduction

For many years, it has been a major concern to increase the N use efficiency in horticultural systems as growing of vegetables often leaves much N in the soil after harvest. This N may leach to deeper soil layers in the form of nitrate, and eutrophy the water environment (Di & Cameron, 2002). This concern is gaining new attention in, e.g. the European Union with the Water Framework Directive (Directive 2000/60/EC), and further demands on horticultural production with the aim to reduce nitrate leaching are expected. Nitrate leaching may be reduced by use of knowledge about root depth of crops (Kristensen and Thorup-Kristensen, 2004a,b; Thorup-Kristensen, 2006a). Deep-rooted crops (>1-m depth) should be grown where much nitrate is available in deep soil layers to increase uptake and

Correspondence: H. L. Kristensen. E-mail: hanne.kristensen@agrsci.dk Received October 2006; accepted after revision June 2007 recycling of N from depth. Thus, root depth should where feasible match the depth distribution of soil N_{inorg} in the design of crop rotations (Thorup-Kristensen, 2006a). On the other hand, it is well known that N distribution may affect root growth locally in the root system, with consequences for N uptake and plant performance (e.g. Forde & Lorenzo, 2001). To enable design of crop rotations with N uptake from deep soil layers it is therefore important to know how N distribution affects root growth and N uptake of crops at field scale.

Knowledge has been gained in laboratory studies about effects of N enrichment in patches on root proliferation, morphology and N uptake (e.g. Drew & Saker, 1975; Hodge, 2004). In enriched patches, root density was found to be higher, and this may be important for inter-species competition on soil N. However, field studies of effects of N distribution on crop root growth are scarce (Schenk *et al.*, 1991; Thorup-Kristensen, 1993; Gao *et al.*, 1998; Christiansen *et al.*, 2006). This is contrasted by numerous studies of

effects of N fertilization on root growth. A review of the studies showed, that restricted N supply or deep placement of N increased density of roots in upper parts of the rooting zone, or in the entire root zone of some crops compared with a large N supply or shallow placement (Schenk et al., 1991; Thorup-Kristensen, 1993; Thorup-Kristensen & Van den Boogaard, 1999). This indicates that moderate N deficiency in crops may result in allocation of resources to the roots that increase the exploration of the soil for nutrients. Other crops showed no effect of N fertilization or distribution on root density and/or root depth (Schenk et al., 1991; Thorup-Kristensen & Van den Boogaard, 1999; Rather et al., 2000; Vamerali et al., 2003: Christiansen et al., 2006). This was contrasted by many studies showing that application of N or organic matter increased root density in surface soil layers (Opena & Porter, 1999; Sharifi et al., 2005) or at depth (Vamerali et al., 2003), and that final root depth was increased (Barraclough et al., 1989). However, in these fertilizer studies, interpretation of the effect of soil Ninorg distribution on root growth cannot be distinguished from the effect of increased availability of nutrients on overall crop performance. It was typical in these studies for the effects on root growth to be connected to large differences in biomass production (e.g. Vamerali et al., 2003). Further due to limited sampling depth, effects of N fertilizer or distribution on maximum root depth and distribution are seldom reported, even for shallow-rooted crops (e.g. Sharifi et al., 2005). Against this background, the question arises of how differences in distribution of soil N_{inorg} affects root density and root depth of crops under conditions where crop N requirements are met, but not greatly exceeded; i.e. under conditions of horticultural production that are efficient in N use. Does deep placement of Ninorg stimulate roots to grow deeper and denser, or is N distribution of minor importance for overall development of root systems?

The distribution of N_{inorg} in soil is affected by a number of factors; one of them is the management of cover crops (green manures and catch crops). Cover crops take up N_{inorg} within their rooting zone. This N is later made available for crop uptake or leaching when incorporated in the topsoil (Thorup-Kristensen *et al.*, 2003; Thorup-Kristensen, 2006b). The time of incorporation of cover crops relative to the periods of high leaching intensity is a major factor that determines the vertical distribution of soil N_{inorg} for the following crop. Thus, different distributions of soil N_{inorg} may be obtained by varying the management of cover crops.

Horticultural production is often characterized by the growing of a large diversity of crops. To design horticultural crop rotations with increased N use efficiency, we need more knowledge about root growth in a wider range of commercial crops. The objectives of this study were to investigate: (a) root distribution and depth in a variety of well-grown vegetable crops; and (b) the effects of deep placement of soil

N_{inorg} on root growth and N uptake of the crops compared with shallow placement.

Materials and methods

Field site and experimental design

Two field experiments were performed with different management of cover crops to obtain different vertical distributions of soil Ninorg. The cover crops were followed by growing a variety of vegetable crops at the Research Centre at Aarslev (10°27'E, 55°18'N) in Denmark. In experiment A, three vegetable crops were grown after two catch crops or bare soil. In experiment B, three vegetable crops were grown after green manure which had been incorporated in the soil in the previous autumn or spring. The soil was a sandy loam (Typic Agrudalf) with the 0- to 0.5-m soil layer containing 1.1% C. 13% clay. 15% silt and 70% sand: the 0.5- to 1-m layer containing 0.3% C, 19% clay, 13% silt and 68% sand and the 1- to 2.5-m layer containing 0.2% C, 18% clay, 14% silt and 68% sand. The pH_{CaCl2} was 6.3, 5.4 and 6.8 in the 0- to 0.5-, 0.5- to 1- and 1- to 2.5-m layers, respectively. Average annual precipitation (624 mm) and air temperature (7.8 °C) were recorded at a meteorological station at the research centre. In both experiments, the vegetables were irrigated when precipitation deficits exceeded 30 mm. The field had been managed according to organic farming practice since 1996.

In experiment A, 16 plots each of $7.5 \times 10 \text{ m}^2$ were arranged in a randomized complete block design with two replicates. Six plots were left bare, six plots were sown with the catch crop Italian ryegrass (Lolium multiflorum Lam. 'Bofur') and four plots with the catch crop fodder radish (Raphanus sativus L. var. oleiformis Pers.) on 8 August 2000 and 31 July 2001. The catch crops were grown after incorporation of a mixture of clover and grass by ploughing. The following spring, the catch crops were incorporated by ploughing on 28 March 2001 and 2 April 2002. Twelveweek-old transplants of leek (Allium porrum 'Parton') were planted after bare soil, ryegrass and radish on 30 May 2001 and 29 May 2002. Beetroot (Beta vulgaris L. ssp. vulgaris var. conditiva Alef. 'Forona') was sown after bare soil and ryegrass on 24 May 2001 and 28 May 2002. Seven and 10week-old transplants of white cabbage (Brassica oleracea L. convar. capitata (L.) Alef. var. alba DC 'Impala') were planted on 10 May 2001 and 27 May 2002, respectively, after bare soil, ryegrass and radish. The distance between rows was 0.5 m, and between plants 0.08 m for leek and 0.5 m for cabbage. Dried chicken manure at 2000 kg ha⁻¹ containing 100 kg N ha⁻¹, was applied to the vegetables on 7 June 2001 and 24 May 2002.

In experiment B, a mixture of clover and grass was undersown in spring barley in spring. The mixture consisted of white clover (*Trifolium repens* L.), black medick (*Medicago*

lupulina L.), birdsfoot trefoil (Lotus corniculatus L.), ryegrass (Lolium perenne L.), timothy (Phleum pratense L.) and meadow fescue (Festuca pratensis Huds.). The green manure was left growing after harvest of the barley in early August and ploughed in autumn (25 October 2001 and 15 November 2002) or spring (2 April 2002 and 19 March 2003). Three vegetables were planted after each green manure treatment in plots arranged in a randomized complete block design with three replicates making a total of 18 plots. An early variety of potato (Solanum tuberosum L. 'Ditta') was planted in $10 \times 3 \text{ m}^2$ plots on 14 May 2002 and 8 May 2003 with 0.75 m between rows and 0.3 m between plants. Four-weekold transplants of Chinese cabbage (Brassica campestris L. ssp. *pekinensis* 'Kasumi') were planted in $10 \times 2.5 \text{ m}^2$ plots on 14 May 2002 and 25 April 2003 with a distance of 0.5×0.5 m, and covered by polypropylene fleece to prevent insect damage. Three- and four-week-old transplants of summer squash (Cucurbita pepo Duch. 'Ambassador') were planted in $10 \times 4.5 \text{ m}^2$ plots on 22 May 2002 and 28 May 2003, respectively, with 1 m between rows and 0.5 m between plants.

Root measurements

Root distribution was measured using minirhizotrons (Kristensen & Thorup-Kristensen, 2004a) which were glass tubes of 1.5 or 3-m length and 70-mm outer diameter. These were inserted along plant rows at an angle of 30° from the vertical at the start of growth. The tubes reached a depth of approximately 1.2 (leek 2001, 2002; potato 2002, 2003 and summer squash 2002) or 2.42 m (all others) into the soil. In experiment A, three minirhizotrons were installed at 0.1-m distance from the rows in each plot. In experiment B, two minirhizotrons were installed, one in and one between the rows, in each plot. This resulted in a total of six replicate minirhizotrons in each cover crop-crop combination in both experiments. Along each minirhizotron, two replicate counting grids (rows of $40 \times 40 \text{ mm}^2$) were painted. A mini-video camera was used to record the roots at the minirhizotron surface. From those recordings, root depth, root intensity and root frequency were obtained. Root depth was measured as the deepest root observed in each of the two counting grids. Root intensity, as an expression of root density, was calculated from the total number of roots crossing the lines in each $40 \times 40 \text{ mm}^2$ as root intersections per metre grid line (intersections m^{-1}). The root frequency, which expresses the ability for depletion of soil Ninorg, was obtained by observing presence/absence of roots crossing the lines in each 40×40 mm². Then the percentage of 40×40 mm² with roots present was calculated for each 0.5-m soil layer (Thorup-Kristensen, 2001). A root depth penetration rate (mm day⁻¹ °C⁻¹) was calculated as the slope of the regression line of the average root depth versus accumulated

daily average temperature from planting or sowing (base temperature of 0 $^{\circ}$ C).

Plant and soil sampling and analysis

Above ground plant biomass was sampled from subplots of 2.25-4.5 m² at harvest in both experiments: leek and white cabbage on 29 October 2001 and 1 October 2002, beetroot including storage root on 24 October 2001 and 1 October 2002, potato including tubers on 22 July 2002 and 17 July 2003, Chinese cabbage on 22 July 2002 and 30 June 2003, and summer squash (>0.25 kg) every week until mid-September including the above ground biomass on 22 July 2002 and 10 September 2003. In both experiments, the samples were kept at 1 °C, chopped, and dried at 80 °C for 20 h. Nitrogen content was determined by a combustion method (thermal conductivity detector, vario-MAX CNS). Soil was sampled for an analysis of Ninorg content to 2.5- and 1.5-m depth in experiments A and B, respectively. In both experiments, nine samples were taken with a soil auger in each plot. The samples were divided into depth intervals of 0.5 m, and pooled to one composite sample for each interval and plot. The composite samples were mixed and a subsample was immediately frozen at -18 °C for later analyses. The frozen soil samples were thawed and 100 g of soil was immediately weighed and extracted in 1 M KCl for 1 h (soil:solution ratio 1:2). The soil extract was filtered and analysed for NH₄⁺ and NO₃⁻ content by flow injection (Lachat 8000). Soil Ninorg content was calculated on an area basis by use of the average volumetric soil density determined for each depth interval.

Statistical analysis

Statistical significance of differences in root distribution, plant and soil pools between crops and treatments was tested by analysis of variance (*F* test). Multiple comparisons were based on values of the least significant difference (LSD) derived from analyses of variance (Proc GLM, SAS Institute Inc., Cary, NC, USA). Root frequencies were transformed prior to analyses by the logit transformation (logitp = $\log[p/(1 - p)]$) to obtain homogeneity of variance. Relationships between root depth and accumulated daily temperature were investigated by simple linear regression modelling and test of homogeneity of slopes. In assessing differences between results, tests with P < 0.05 were considered statistically significant. Annual differences were small relative to differences between crops and treatments, and results are therefore presented as averages over the two experimental years.

Results

In both experiments, the management of cover crops affected the soil N_{inorg} distribution in spring, and the management of cover crop followed by the growing of

crops affected the soil N_{inorg} at harvest (Figures 1 and 2). In experiment A in spring, significant differences were seen in the 0- 0.5-m soil layer between bare soil, ryegrass and fodder radish; and in the 0.5- to 2.5-m soil layer between bare soil and rye grass/fodder radish (Figure 1a). No differences were found in the total amount of soil Ninorg in the 0- to 2.5-m soil layer between cover crop managements in spring (180 kg N ha⁻¹). At harvest of the vegetable crops in experiment A, differences of up to 134 kg N ha⁻¹ were recorded between the total amounts of Ninorg left on average below crops (Table 1). Differences for the same species were up to 73 kg N ha⁻¹ depending on cover crop management. The differences were found mainly in the soil layers below 0.5-m depth (Figure 1b). The average N content in the above ground biomass differed by up to 107 kg ha⁻¹ among crops in experiment A (Table 1). A difference of 38 kg ha⁻¹ was found in the N content of leek depending on cover crop management. The vields of saleable product of leek, beetroot and white cabbage amounted to 34, 49 and 63 t fresh weight ha⁻¹, respectively.



Figure 1 Distribution of soil inorganic N in soil layers in experiment A, (a) in spring after incorporation of a cover crop of bare soil (BS), ryegrass (RG) or fodder radish (FR); and (b) at harvest of a crop of leek, beetroot or white cabbage grown after the cover crops. Bars indicate standard errors for averages over 2 years (n = 2).

In experiment A, root depth differed at harvest between species by up to 1.9 m (Table 2, Figure 3), and the rate of root depth penetration (slope of line from linear regression) differed between species by up to 1.22 mm day⁻¹ °C⁻¹. Within species, differences were found for white cabbage of 0.14 mm day⁻¹ °C⁻¹ when grown after bare soil instead of after cover crops. Root intensity of the vegetable crops in experiment A differed strongly among species in the 0.5- to 2-m layer (Figure 4). Differences within species were found for white cabbage with higher intensities in the 0.5- to 1-m layer; and lower intensities in the 1.5- to 2-m layer when grown after a cover crop, compared with bare soil. The root frequency was very low for all three crops in June in experiment A (Figure 5). In July, frequencies were still below 30% and few differences between species were found. In August, frequencies were much larger and there were clear differences between species. Root frequencies were much larger in September, especially in the 0- to 0.5-m layer for leek and in the 0.5- to 2-m layer for beetroot and white cabbage. In October, differences between species were still pronounced but only in layers below 0.5-m depth. At this time the root frequency reached the maximum value for leek in the 0- to 0.5-m layer, and frequencies of beetroot and white cabbage had increased in the 1- to 2.4-m layer. Differences within species were seen for beetroot in the 0- to 0.25-m layer in October 2002. For white cabbage, root frequency was greater in the 0.5- to 0.75-m layer when cabbage was grown after fodder radish instead of after ryegrass/bare soil (August 2001) or ryegrass (September 2001). In the deeper soil layers, frequencies were the greatest when white cabbage was grown after bare soil compared with after fodder radish in the 1.25to 1.5-m layer (August 2002) and with fodder radish/ryegrass in the 1.5- to 2.4-m layer (October 2001).

In experiment B in spring, the concentrations of soil N_{inorg} in the 0.5- to 1.5-m layer was larger after incorporation of cover crops in the autumn compared with after incorporation in the spring (Figure 2a). No differences were found in the total amount of soil Ninorg in the 0- to 1.5-m soil layer between cover crop managements in spring (167 kg N ha^{-1}). At harvest of the vegetable crops in experiment B, differences of up to 38 kg N ha⁻¹, on average, were recorded between total amounts of soil N_{inorg} remaining at harvest (Table 1). Within species, differences were up to 20 kg N ha⁻¹. The concentrations were largest in the soil layers below potato (Figure 2b). The average N content in the above ground biomass differed by up to 106 kg ha⁻¹ between species in experiment B (Table 1). Differences within species were found for N content in potato and summer squash of 28 and 32 kg ha⁻¹, respectively. The yields of saleable product of potato, Chinese cabbage and summer squash amounted to 29, 55 and 157 t fresh weight ha^{-1} , respectively.

In experiment B, root depth at harvest differed between species by up to 1.2 m (Table 2, Figure 3). Also the rate of root depth penetration differed between species by up to

Main crop	Cover crop management	Soil N _{inorg} placement	Plant biomass (t dry weight ha ⁻¹)	Plant N $(kg ha^{-1})$	Soil inorganic N at harvest (kg ha ⁻¹)	
Experiment A						
Leek	Bare soil	Deep	7.52a	113a	176a*	
	Ryegrass	Shallow	7.35a	130ab	153a*	
	Radish	Shallow	8.17a	151bc	176a*	
Beetroot	Bare soil	Deep	9.91b	170cd	153a*	
	Ryegrass	Shallow	10.16b	193d	80b*	
White cabbage	Bare soil	Deep	11.61c	251e	42bc*	
	Ryegrass	Shallow	11.43c	236e	35c*	
	Radish	Shallow	11.37c	229e	26c*	
Experiment B						
Potato	Autumn incorporation	Deep	8.21b	98a	$90a^{\dagger}$	
	Spring incorporation	Shallow	9.02b	126b	$70b^{\dagger}$	
Chinese cabbage	Autumn incorporation	Deep	4.15a	97a	$44cd^{\dagger}$	
	Spring incorporation	Shallow	4.37a	102ab	40cd^{\dagger}	
Summer squash	Autumn incorporation	Deep	10.53c	189c	53c [†]	
	Spring incorporation	Shallow	10.90c	221d	35d [†]	

Table 1 Plant biomass, N content and soil inorganic N remaining at harvest in experiments A and B

Numbers followed by different letters are significantly different within each experiment (P = 0.05). *Soil inorganic N measured in the 0- to 2.5-m soil layer in experiment A. [†]Soil inorganic N measured in the 0- to 1.5-m soil layer in experiment B.

Table 2 Root depth at harvest and outputs from a linear regression analysis of root depth (*Y*) versus accumulated daily temperature above 0 °C (*X*) in experiments A and B (Figure 2)

Main crop	Cover crop management	Soil N _{inorg} placement	Root depth at harvest (m)	Output from linear regression		
				Y intercept	Slope (mm day ⁻¹ °C ⁻¹)	r^2
Experiment A						
Leek	Bare soil	Deep	0.48a	0.02	0.23a	0.90
	Ryegrass	Shallow	0.50a	-0.02	0.25a	0.95
	Radish	Shallow	0.49a	0.00	0.24a	0.89
Beetroot	Bare soil	Deep	1.83b	-0.48	1.00b	0.93
	Ryegrass	Shallow	1.87b	-0.47	1.02b	0.94
White cabbage	Bare soil	Deep	>2.42c	-0.66	1.55d	0.97
	Ryegrass	Shallow	>2.42c	-0.64	1.36c	0.93
	Radish	Shallow	>2.42c	-0.74	1.47c	0.98
Experiment B						
Potato	Autumn incorporation	Deep	0.75a	-0.38	0.90a	0.86
	Spring incorporation	Shallow	0.60a	-0.26	0.72a	0.73
Chinese cabbage	Autumn incorporation	Deep	1.44b	-0.44	1.56c	0.92
	Spring incorporation	Shallow	1.23c	-0.27	1.17b	0.91
Summer squash	Autumn incorporation	Deep	2.02e*	-0.34	1.34c [†]	0.99
	Spring incorporation	Shallow	1.73d*	-0.08	$1.06b^{\dagger}$	0.91

Numbers followed by different letters are significantly different within each experiment (P = 0.05). *Root depth at harvest from 1 year only (2003). [†]The slopes presented for summer squash are based on the results from 2002 and 2003, but the difference between autumn and spring incorporation was statistically significant only in 2003.

 $0.56 \text{ mm day}^{-1} \circ \text{C}^{-1}$ except between Chinese cabbage and summer squash. Differences within species were found for Chinese cabbage and summer squash of 0.21 and 0.29 m for root depth at harvest, and 0.39 and 0.28 mm day⁻¹ $\circ \text{C}^{-1}$,

respectively, for root depth penetration rate. Root frequencies were generally greater in July in experiment B (Figure 6) than in July in experiment A (Figure 5). In experiment B, the frequencies were much greater in August than in July



Figure 2 Distribution of soil inorganic N in soil layers in experiment B, (a) in spring after incorporation of a cover crop in the previous autumn (AI) or spring (SI); and (b) at harvest of a crop of potato, Chinese cabbage or summer squash grown after the cover crops. Bars indicate standard errors for averages over 2 years (n = 2). Results for summer squash at harvest are from 2003 only.

especially for summer squash; and differences were found between potato and summer squash. Chinese cabbage had already been harvested at this time and therefore no data were recorded. In September, frequencies had increased strongly in the 1- to 2-m soil layer under summer squash compared with the previous measurement. Differences within species were found for Chinese cabbage in July in the 1- to 1.5-m layer and for summer squash in September 2003 in the 1.5- to 2-m layer.

Figure 7 shows the relationship between root frequency and the relative decrease in soil N_{inorg} from spring until harvest in each soil layer in both experiments. The results show that at root frequencies below 5% there was no decrease in soil N_{inorg} , and at frequencies above 20%, N_{inorg} was decreased by 40–100%. Potato and beetroot were exceptions to this relationship with frequencies of between 20 and 40% resulting in decreases in Soil N_{inorg} of 20% or less (Figure 7: potato layer 2, beetroot layer 4).

Discussion

Root growth of species

All crops continued root growth until harvest. Nevertheless, the vertical distribution of roots and the root depth varied



Figure 3 Rooting depth versus accumulated daily temperature above 0 °C from sowing or planting in experiment A: (a) leek, beetroot and white cabbage after a previous crop of bare soil (BS), ryegrass (RG) or fodder radish (FR); and in experiment B: (b) potato, Chinese cabbage and summer squash after cover crop incorporation in the previous autumn (AI) or spring (SI). Each symbol represents a point in time in 1 year. Outputs from a linear regression analysis are shown in Table 1.



Figure 4 Root intensity at harvest in early September in experiment A of leek, beetroot and white cabbage after a previous crop of bare soil (BS), ryegrass (RG) or fodder radish (FR). Bars indicate standard errors for averages over 2 years (n = 2).



widely between species at harvest, and the rate of root depth penetration varied even more. For the deep-rooted crops, root growth from August until harvest was restricted entirely to below 0.75-m depth as expressed by the root frequency. The results on root growth of Chinese cabbage are, to our knowledge, the first reported in the literature. The growth season was short, but due to the relatively high rate of depth penetration, Chinese cabbage still reached 1.3-m depth at harvest in July. The rate of root depth penetration was comparable with that of another Brassica, white cabbage, in this and other studies (Kristensen & Thorup-Kristensen, 2004b; Thorup-Kristensen, 2006b). The root frequency was relatively small in the surface soil layer, but comparable with frequencies obtained by white cabbage at a similar accumulated daily temperature. White cabbage was the deepest-rooted crop in the present study due to the long growing season and high rate of depth penetration. The results for beetroot confirmed a root depth of 1.7 m and a penetration rate of 0.9 mm day⁻¹ °C⁻¹ found by Christiansen *et al.* (2006), whereas a higher rate of 1.6 mm day⁻¹ °C⁻¹ was found under semi-field conditions in the Wageningen Rhizolab (Smit & Groenwold, 2005). The closely related crop of sugar beet has been found to reach 2- to 3-m depth in the field (e.g. Weaver & Bruner, 1927; Vamerali et al., 2003). The root intensity and frequency of beetroot were smaller than those of the deep-rooted Brassicas and summer squash in accordance with Smit & Groenwold (2005). Summer squash roots grew very deep like other Cucurbits such as squash and pumpkin with root depths of 2 m (Weaver & Bruner, 1927), and watermelon with at least 1.5 m (Xie et al., 2006). The root depth reached by summer squash was due to the long growing season in combination with a relatively high rate of root depth penetration. The shallow root depth found for leek confirmed previous field results of 0.4-0.6 m (Weaver & Bruner, 1927; Smit et al., 1996). The rate of root depth penetration and proliferation were both slow, resulting in low root intensity at harvest as previously reported for leek in comparison with other vegetable crops (Smit et al., 1996; Smit & Groenwold, 2005). Potato also was found to be shallow rooted as reported in the literature with depths of 0.6-1 m (Parker et al., 1989; Ruijter et al., 1996). The rate of root depth penetration was comparable with that of deeper rooted crops like beetroot and carrot (Thorup-Kristensen & Van den Boogaard, 1999; Kristensen & Thorup-Kristensen, 2004b). Therefore, the shallow root depth was due to the short growing season of this early variety, and to the leaf loss

Figure 5 Root frequency through the growing season in experiment A of leek, beetroot and white cabbage after a previous crop of bare soil (BS), ryegrass (RG) or fodder radish (FR). The accumulated daily temperature above 0 °C from sowing or planting until the recording of root frequency is indicated in the lower right corner. Bars indicate standard errors for averages over 2 years (n = 2).



Figure 6 Root frequency through the growing season in experiment B of potato, Chinese cabbage (at harvest only) and summer squash after previous incorporation of cover crop in autumn (AI) or spring (SI). The accumulated daily temperature above 0 °C from sowing or planting until recording of root frequency is indicated in the lower right corner. Results for summer squash below 1-m depth are from 2003 only. Bars indicate standard errors for averages over 2 years (n = 2).

caused by potato blight as commonly seen in organic farming. This was confirmed by Stalham & Allen (2001) who found that a considerable range in root depths of 0.6–1.4 m was determined by differences in length of growing season and not in rate of root depth penetration in potato cultivars. The root distributions obtained for the six crop species were confirmed by the large reductions of N_{inorg} in the rooted soil layers indicating crop uptake of soil N_{inorg}.



Figure 7 The decrease in soil N_{inorg} from spring until harvest as a percentage of soil N_{inorg} in spring versus root frequency at harvest for each 0.5-m soil layer in experiment A of leek, beetroot and white cabbage; and in experiment B of potato, Chinese cabbage and summer squash. Each symbol represents an average over 2 years. The subscript number indicates the depth of the soil layer: 1 = 0-0.5 m; 2 = 0.5-1 m; 3 = 1-1.5 m; 4 = 1.5-2 m and 5 = 2-2.5 m. Soil layers with N_{inorg} content < 1.8 mg N kg⁻¹ soil in spring or without roots at harvest were excluded.

Effects of N distribution on root growth

Three of the four deep-rooted crops in the two experiments, white cabbage, Chinese cabbage and summer squash, responded to more N_{inorg} at depths. This was also found in a study of broccoli to 1-m depth, where root intensity and depth were increased by deep placement of N_{inorg} compared with shallow placement (Thorup-Kristensen, 1993). Also wheat had greater root density in the 0.3- to 0.9-m soil layer when nitrate had leached to this depth from the surface soil (Gao et al., 1998). This is in contrast to studies of two Brassicas, kohlrabi and cauliflower, showing effects on yields but not on root density with different vertical distributions of Ninorg or N fertilization (Schenk et al., 1991; Rather et al., 2000). The lack of effect could possibly be due to the limited sampling depth of 0.6 m. On the other hand, a study of carrot to 1-m depth showed no effects on root intensity in the deeper parts of the measuring zone and root depth penetration, but in the upper parts decrease in root intensity and less spread of roots into inter-rows were found. This was a response to larger availability of Ninorg after incorporation of leguminous cover crops compared with smaller availability (Thorup-Kristensen & Van den Boogaard, 1999). Also spinach showed decreased root density in the upper parts of a measuring zone of 0.6-m depth and no effects in the deeper parts as a response to shallow placement of soil Ninorg (large availability) when compared with deep placement (Schenk et al., 1991). For beetroot in the present study, the lack of effect on root growth of N_{inorg} distribution contrasts with the results of Christiansen et al. (2006) obtained on the same

soil: beetroot increased root depth by 0.5 m and root intensity was more than doubled in the 0.5- to 2-m soil layer due to deep placement of Ninorg. This response may be explained by the fact that the amount of soil N_{inorg} was smaller than in the present experiment. However, the biomass production was greater. On the other hand, the closely related sugar beet showed no effect on maximum root depth whereas yield, root growth rate and root density were increased in the 0.3to 1.8-m soil layer due to N fertilization (Vamerali et al., 2003). This indicates that the response of beetroot to N_{inorg} distribution depends on the N status of the crop in combination with other factors affecting the resource allocation during growth. For the shallow-rooted leek and potato, the shallow placement of Ninorg caused a larger N uptake but root growth was unaffected. Thus, for these shallow-rooted crops the management of cover crops caused differences in Ninorg availability more than in Ninorg distribution, as the deep Ninorg was out of reach of the shallow roots. The results contrast with previous studies of potato where root density and depth were increased due to fertilizer application and subsequent increase in biomass production (Opena & Porter, 1999; Sharifi et al., 2005). However, in spinach, another shallow-rooted crop, limiting N supply or deep placement of N_{inorg} have both been shown to decrease yield, but increase root density in surface soil layers (Schenk et al., 1991). In general, the effect of N_{inorg} distribution on root growth is likely to be closely related to the overall performance and N nutritional status of the crop. In the present study, the basic N needs of the crops were met, in the sense that N availability was adequate to ensure good development and yield of all six crops. Still, the effect of differences in soil Ninore distribution on root growth was found to differ between species and experiments with no clear relationship to the N status of the crop.

Effect of root growth on N uptake

As noted earlier deep placement of soil Ninorg increased the root depth penetration by 0.2- to 0.4-m at harvest for three of the four deep-rooted crops compared with shallow placement. These increases of root depth also increased the availability of N_{inorg} by giving access to a larger volume of soil at depth. A rough calculation, using the concentration of soil Ninorg found in spring at deep Ninorg placement, shows that a 0.2- to 0.4-m increase in rooting depth may give access to 9-28 kg N ha⁻¹. Also the changes of root frequency due to Ninorg distribution may have affected the rate of Ninorg depletion of the soil. This would influence the amount of Ninorg left after harvest especially in the deepest and most recently rooted part of the root zones, where time for N uptake was limited. Large differences between species, were found in the amount of Ninorg left at harvest in the deep soil layers (Table 1, Figures 1b and 2b), and these differences were caused by a combination of the vertical distribution of N_{inorg} in spring and the root depth of the vegetable crop. Thus, deep N_{inorg} placement followed by a shallow-rooted crop left the largest amount, and shallow N_{inorg} placement followed by a deep-rooted crop left the smallest amount. The importance of the crop species for N cycling in crop rotations in experiment A is discussed in Thorup-Kristensen (2006a,b). When comparing the effect of species and vertical distribution of soil N_{inorg} on root distribution and depth, the differences between species were much larger than the differences caused by the vertical distribution of soil N_{inorg} , and the species were much more important than the distribution of N_{inorg} for the capacity to deplete N at depth.

The root frequency was calculated to express the ability for soil Ninorg depletion. The calculation is based on the theory that a few evenly distributed roots are sufficient for uptake of the highly mobile NO_3^- ion, and that these roots therefore can exhaust soil layers for NO3⁻ over the season (Thorup-Kristensen, 2001). The relationship between root frequencies and decrease in soil Ninorg was explored in Figure 7. It showed that soil layers with root frequencies below 5% led to a small decrease in N_{inorg}. These soil layers were from the deepest part of the root zones that had been recently rooted, and therefore the roots had limited time for N_{inorg} uptake. When root frequencies were above 20%, the Ninorg decrease was at least 40%. Exceptions from this were the deep parts of the root zone of potato and beetroot which indicated a low efficiency in N depletion by these species. This was probably because the availability of soil Ninorg exceeded the N demand. For the other species, there was no further relationship between root frequencies and decreases of Ninorg. Thus, the measure of root frequency was of limited value for expression of the ability for Ninorg depletion in the present study. However, the lack of relationship may have been caused by the influence of factors other than crop N uptake on N_{inorg} decrease, such as mineralization of soil organic N, leaching and N application during the experiments.

Conclusion

Deep placement of soil N_{inorg} may increase root depth and density in the deep parts of the rooting zone in some crops compared with shallow placement. The effect may depend on the N status of the crop. However, the differences in root growth caused by vertical distribution of soil N_{inorg} are much smaller than the inherent difference found between species. This means that the capacity of crops to take up N at depth should be based primarily on the inherent rooting depth of species. Crop rotations should be designed to match root distribution with soil N_{inorg} distribution at depth. Effects of vertical distribution of soil N_{inorg} on root growth do not need to be taken into account in the design of crop rotations, as these effects are of minor importance for the ability of root systems to take up soil N_{inorg} at depth.

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