

MYCORRHIZA-MEDIATED DISEASE RESISTANCE - A MINI-REVIEW

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Abstract

Arbuscular mycorrhizal fungi (AMF) play an essential role as one of the primary mutualistic plant-microbe symbioses. Plants benefit from root endophytes that extend their zone of activity beyond the rhizosphere (Feddermann *et al.*, 2010; Hohmann *et al.*, 2011, 2012). The main known benefits of mycorrhiza involve nutrient mobilisation (mainly phosphorus), improved tolerance against abiotic (mainly drought) and biotic stresses (mainly soil-borne pathogens) (Azcón-Aguilar & Barea, 1996; Parniske, 2008). An increasing number of studies highlight a significant role of AMF in the mediation of disease resistances. Besides an improved phosphorus use efficiency, individual reports have shown enhanced levels of defence-related compounds (such as glucanases, chitinases and phenolics) in mycorrhizal plants, and there is first evidence of certain phytohormone pathways (in particular jasmonate signalling) to be involved in mycorrhiza-mediated disease resistance (Jung *et al.*, 2012).

Despite their ecological and nutritional importance, legume crops receive less and less attention in breeding programmes and crop rotations. Increasing problems with fungal diseases seem to be a main cause for the decline in yield (Wilbois, 2011; Finckh *et al.*, 2013). However, legumes are known to show particularly strong interactions with important symbionts such as AMF and *Rhizobium* spp. and, therefore, provide a valuable model system to identify genotypes that interact efficiently with symbiotic microbes.

Linkage mapping was used to identify quantitative trait loci (QTL) and genes in pea that are linked to resistance against various pathogens (Prioul-Gervais *et al.*, 2007; Pereira *et al.*, 2009; Fondevilla *et al.*, 2011; Hamon *et al.*, 2011; Li *et al.*, 2012). The ability of plants to respond to AMF can vary widely between plant species and among genotypes (Parke & Kaepller, 2000; Sawers *et al.*, 2010). Genotypic differences in the response to AMF have been observed in various crops (Powell *et al.*, 1982; Hetrick *et al.*, 1993; Kaepller *et al.*, 2000; Tawaraya *et al.*, 2001). Such differences in mycorrhizal responsiveness indicate a genetic basis for plant-AMF interactions. Galván *et al.* (2011) were the first to report on QTL governing responses of onion species to AMF based on shoot biomass. However, little is known about the genetic basis for mycorrhiza-mediated disease resistance and more research is needed to exploit genotypic differences, e.g., via marker-assisted selection.

For legume crops, the use of association mapping has been restricted in the past due to insufficient genome-wide marker coverage. Most recently, 384 pea accessions of the United States Department of Agriculture – Agricultural Research Service (USDA-ARS) pea core collection were genotyped for 20.000+ single nucleotide polymorphism (SNP) marker using genotyping-by-sequencing. These SNP markers are currently being mapped *in silico* using *M. truncatula* to establish a physical genetic map. Further, a linkage map of a portion of these SNP markers will also soon be created using a pea recombinant inbred population. This high density genotyped association mapping panel can now be used for genome-wide association studies as demonstrated for barley (Pasam *et al.*, 2012; Shu *et al.*, 2012).

References

- Azcón-Aguilar C** & Barea J (1996) Arbuscular mycorrhizas and biological control of soil-borne plant pathogens – an overview of the mechanisms involved. *Mycorrhiza* **6**: 457–464.
- Feddermann N**, Finlay R, Boller T & Elfstrand M (2010) Functional diversity in arbuscular mycorrhiza – the role of gene expression, phosphorous nutrition and symbiotic efficiency. *Fungal Ecol* **3**: 1–8.
- Finckh MR**, Bruns C, Demmel M, et al. (2013) Soil and plant health management for improved sustainability in organic grain legume production. *Sustainability* **5**: 1-37.
- Fondevilla S**, Almeida NF, Satovic Z, Rubiales D, Vaz Patto MC, Cubero JI & Torres AM (2011) Identification of common genomic regions controlling resistance to *Mycosphaerella pinodes*, earliness and architectural traits in different pea genetic backgrounds. *Euphytica* **182**: 43–52.
- Galván G**, Kuyper TW, Burger K, Keizer LCP, Hoekstra RF, Kik C & Scholten OE (2011) Genetic analysis of the interaction between *Allium* species and arbuscular mycorrhizal fungi. *Theor Appl Genet* **122**: 947–960.
- Hamon C**, Baranger A, Coyne CJ, et al. (2011) New consistent QTL in pea associated with partial resistance to *Aphanomyces euteiches* in multiple French and American environments. *Theor Appl Genet* **123**: 261–281.
- Hetrick BAD**, Wilson GWT & Cox TS (1993) Mycorrhizal dependence of modern wheat cultivars and ancestors: a synthesis. *Can J Bot* **71**: 512–518.
- Hohmann P**, Jones EE, Hill RA & Stewart A (2011) Understanding *Trichoderma* in the root system of *Pinus radiata*: associations between rhizosphere colonisation and growth promotion for commercially grown seedlings. *Fungal Biol* **115**: 759–767.
- Hohmann P**, Jones EE, Hill RA & Stewart A (2012) Ecological studies of the bio-inoculant *Trichoderma hamatum* LU592 in the root system of *Pinus radiata*. *FEMS Microbiol Ecol* **80**: 709–721.
- Jung SC**, Martinez-Medina A, Lopez-Raez J a & Pozo MJ (2012) Mycorrhiza-induced resistance and priming of plant defenses. *J Chem Ecol* **38**: 651–664.
- Kaeppler SM**, Parke JL, Mueller SM, Senior L, Stuber C & Tracy WF (2000) Variation among Maize Inbred Lines and Detection of Quantitative Trait Loci for Growth at Low Phosphorus and Responsiveness to Arbuscular Mycorrhizal Fungi. *Crop Sci* **40**: 358.
- Li W**, Feng J, Chang K, Conner R, Hwang S, Strelkov S, Gossen B & McLaren D (2012) Microsatellite DNA markers indicate quantitative trait loci controlling resistance to pea root rot caused by *Fusarium avenaceum*. *Plant Pathol J* **11**: 114–119.
- Parke J** & Kaeppler S (2000) Effects of genetic differences among crop species and cultivars upon the arbuscular mycorrhizal symbiosis. *Arbuscular mycorrhizas: physiology and function*, (Kapulnik Y & Douds DD, eds), pp. 131–146. Springer Netherlands, Kluwer, Dordrecht
- Parniske M** (2008) Arbuscular mycorrhiza: the mother of plant root endosymbioses. *Nat Rev Microbiol* **6**: 763–775.
- Pasam RK**, Sharma R, Malosetti M, van Eeuwijk F a, Haseneyer G, Kilian B & Graner A (2012) Genome-wide association studies for agronomical traits in a world wide spring barley collection. *BMC Plant Biol* **12**: 16.
- Pereira G**, Marques C, Ribeiro R, Formiga S, Dâmaso M, Tavares Sousa M, Farinhó M & Leitão JM (2009) Identification of DNA markers linked to an induced mutated gene conferring resistance to powdery mildew in pea (*Pisum sativum* L.). *Euphytica* **171**: 327–335.
- Powell CL**, Clark GE & Verberne NL (1982) Growth response of four onion cultivars to several isolates of VA mycorrhizal fungi. *New Zeal J Agric Res* **25**: 465–470.
- Prioul-Gervais S**, Deniot G, Receveur E-M, Frankewitz A, Fourmann M, Rameau C, Pilet-Nayel M-L & Baranger A (2007) Candidate genes for quantitative resistance to *Mycosphaerella pinodes* in pea (*Pisum sativum* L.). *Theor Appl Genet* **114**: 971–984.
- Sawers RJH**, Gebreselassie MN, Janos DP & Paszkowski U (2010) Characterizing variation in mycorrhiza effect among diverse plant varieties. *Theor Appl Genet* **120**: 1029–1039.
- Shu X**, Backes G & Rasmussen SK (2012) Genome-wide Association Study of Resistant Starch (RS) Phenotypes in a Barley Variety Collection. *J Agric Food Chem* **60**: 10302–10311.
- Tawaraya K**, Tokairin K & Wagatsuma T (2001) Dependence of *Allium fistulosum* cultivars on the arbuscular mycorrhizal fungus *Glomus fasciculatum*. *Appl Soil Ecol* **17**: 119–124.
- Wilbois K-P** (2011) Steigerung der Wertschöpfung ökologisch angebauter Marktfrüchte durch Optimierung des Managements der Bodenfrucht-barkeit. FiBL report.